



THE UNIVERSITY *of* EDINBURGH

Edinburgh Research Explorer

A rare de novo nonsense mutation in OTX2 causes early onset retinal dystrophy and pituitary dysfunction

Citation for published version:

Henderson, RH, Williamson, KA, Kennedy, JS, Webster, AR, Holder, GE, Robson, AG, FitzPatrick, DR, van Heyningen, V & Moore, AT 2009, 'A rare de novo nonsense mutation in OTX2 causes early onset retinal dystrophy and pituitary dysfunction' *Molecular Vision*, vol 15, no. 260, pp. 2442-7.

Link:

[Link to publication record in Edinburgh Research Explorer](#)

Document Version:

Publisher's PDF, also known as Version of record

Published In:

Molecular Vision

General rights

Copyright for the publications made accessible via the Edinburgh Research Explorer is retained by the author(s) and / or other copyright owners and it is a condition of accessing these publications that users recognise and abide by the legal requirements associated with these rights.

Take down policy

The University of Edinburgh has made every reasonable effort to ensure that Edinburgh Research Explorer content complies with UK legislation. If you believe that the public display of this file breaches copyright please contact openaccess@ed.ac.uk providing details, and we will remove access to the work immediately and investigate your claim.



A rare de novo nonsense mutation in *OTX2* causes early onset retinal dystrophy and pituitary dysfunction

Robert H. Henderson,^{1,2} Kathleen A. Williamson,³ Joanna S. Kennedy,³ Andrew R. Webster,^{1,2}
Graham E. Holder,^{1,2} Anthony G. Robson,^{1,2} David R. FitzPatrick,³ Veronica van Heyningen,³
Anthony T. Moore^{1,2}

¹Moorfields Eye Hospital, London, United Kingdom; ²UCL Institute of Ophthalmology, London, United Kingdom; ³MRC Human Genetics Unit, Western General Hospital, Edinburgh, Scotland

Purpose: To describe the clinical findings of a patient with an early onset retinal dystrophy and a novel mutation in *OTX2*, and to compare these findings with previously reported cases.

Methods: Using direct sequencing, we screened 142 patients, who had either Leber congenital amaurosis (LCA) or early onset retinal dystrophy (EORD), for mutations in *OTX2*. All patients received a detailed ophthalmic examination including electroretinography and retinal imaging.

Results: Only one mutation in *OTX2* was identified. A novel heterozygous p.S138X stop mutation was identified in a seven-year-old male who had an infantile onset retinal dystrophy. The mutation was not present in either parent or in 181 blood donor samples. There was a history of failure to thrive in infancy, poor feeding, and growth hormone deficiency. Poor vision and nyctalopia was present from the first year. Funduscopy revealed a hyperpigmented peripapillary ring with a fine granular pigmentation of the RPE throughout the fundus. The scotopic bright flash ERG a-wave was subnormal and the waveform electronegative, in keeping with dysfunction both at the level of the photoreceptor and post-phototransduction. Visual function has been stable to date.

Conclusions: Mutations in *OTX2* have been reported in association with major developmental malformations of the eye, with retinal dystrophies such as LCA, and with pituitary dysfunction and seizure activity in some cases. This case adds further support for a role of *OTX2* both in retinal development and pituitary function, and highlights a novel retinal dystrophy phenotype seen in association with mutations in *OTX2*.

The orthodenticle protein homolog 2 (*OTX2*) is a homeobox gene that plays a critical role in retinal photoreceptor development. The gene, on chromosome 14q23.1, is organized into five exons, only three of which are translated. It contains a highly conserved homeodomain. *OTX2* is expressed during development in the neuroepithelium of most of the forebrain and midbrain, including the eye domain [1]. Complete elimination of *OTX2* function in mice by gene targeting results in the absence of the forebrain and embryonic lethality [2,3]. While *Otx2* null embryos display a severe cranial phenotype, lacking the anterior neuroectoderm, and with abnormalities in body plan [2,3], heterozygous deletion of *Otx2* leads to a variable phenotype that is dependent on genetic background. *Otx2*^{+/-} mice can be normal, have developmental eye anomalies including anterior segment malformations, severe eye abnormalities such as microphthalmia or anophthalmia, or head abnormalities [2,4]. In early development *Otx2*, specifies the anterior neuroectoderm during gastrulation [2] and is expressed throughout the forebrain and midbrain in the developing embryo [1]. Once eye development is specified,

expression of *Otx2* narrows. It is required for the development of the presumptive retinal pigment epithelium (RPE) [5], where it has a continuing maintenance role throughout adulthood [6]. Analysis of *Otx2*^{-/-} mice has shown that it is essential for specification of the RPE; loss of *OTX2* function results in the outer layer of the optic cup developing into ectopic neural retina [7]. However, *OTX2* is also required for the development and maintenance of the neural retina. *Otx2* protein is found in bipolar and in ganglion cells [6,8,9], while in the outer nuclear layer it regulates the expression of the closely related *Crx* gene that controls the expression of a suite of photoreceptor function genes, including opsins. *Otx2* and *Crx* can also bind to common DNA-binding sequences [10]. Ablation of *OTX2* in retinal progenitor cells has been shown to affect RPE development and leads to a significant decrease in rod photoreceptors and bipolar cells, suggesting that *OTX2* is also involved in retinal bipolar cell development [8]. It has recently been demonstrated that *OTX2* protein is transported from the retina to the visual cortex in the developing rat, and appears to modify postnatal synaptic plasticity [9].

OTX2 mutations in humans have been associated with a range of ophthalmological phenotypes including anophthalmia, microphthalmia, developmental anomalies of the optic nerve and chiasm, and Leber congenital amaurosis (LCA). As *OTX2* plays a key role in retinal development, and

Correspondence to: A.T. Moore, Moorfields Eye Hospital, 162 City Road, London, EC1V 2PD, United Kingdom; Phone: +44 207 566 2260; FAX: +44 207 608 6830; email: tony.moore@ucl.ac.uk

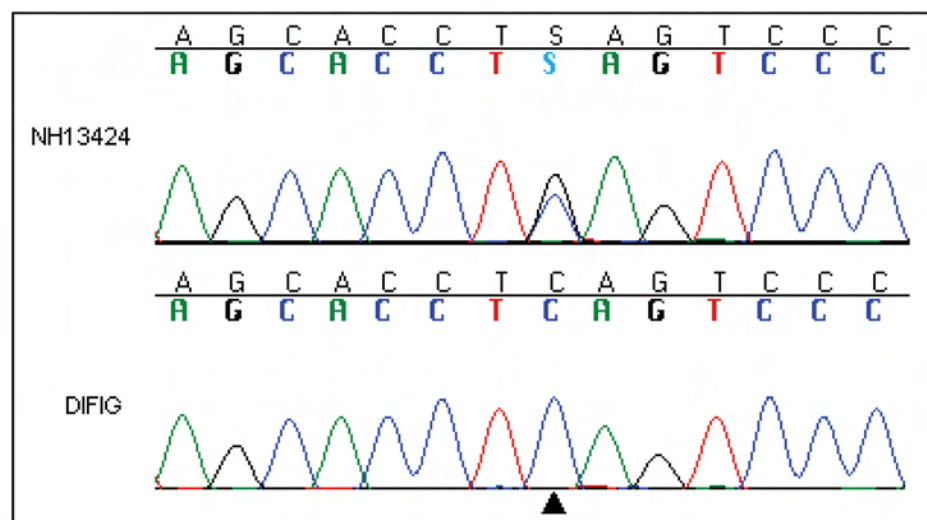


Figure 1. Direct sequencing. At the position indicated by the black arrowhead, the heterozygous C>G change is revealed in the DNA sequence trace for patient NH13424. Below this the wild type trace for normal control subject DIFIG is shown.

a mutation in *OTX2* has been reported in association with an infantile onset retinal dystrophy [11], we screened a large panel of patients with LCA and other forms of childhood onset retinal dystrophy for mutations in this gene.

METHODS

A panel of DNA from 142 patients with LCA and severe childhood onset retinal dystrophy were recruited at Moorfields Eye Hospital and The Hospital for Children, Great Ormond Street, London, as part of an ongoing molecular genetic study of childhood retinal dystrophies. The research has local research ethics committee approval and conformed to the tenets of the Declaration of Helsinki. A full clinical history was taken and examination performed in all subjects. In older children and adults this included, where possible, best corrected logMAR visual acuity and color vision (Hardy Rand Rittler, Richmond Products, Albuquerque, NM); slit-lamp biomicroscopy or indirect ophthalmoscopy; Goldmann visual fields; autorefractometer (Luneau L62-3D autorefractometer - LUNEAU Chartres, France), axial length measurement, and auto-keratometry (IOL Master, Carl Zeiss Meditec, Dublin, CA); optical coherence tomography (Stratus OCT™ - software version 3; Carl Zeiss Meditec, Inc.); fundus photography (TRC-501X with IMAGEnet 2000 system software—TOPCON Corporation, Japan), and fundus autofluorescence (Heidelberg Retinal Angiograph-II, Heidelberg Engineering, Heidelberg, Germany). All patients had electroretinography performed either at our institutions or at the referring hospital. Most patients had tests performed at diagnosis which, in the older patients, predated the ISCEV standards. Patients seen more recently, or in whom we wished to confirm the diagnosis, had electroretinography conducted in accordance with current International Society for Clinical Electrophysiology of Vision standards using gold foil or DTL electrodes [12,13]. ERG testing in infants and young children

was performed using skin electrodes in accordance with previously published protocols [14,15].

DNA was extracted from whole blood obtained from each affected individual and immediate family members using the Nucleon BACC-2 kit (GE Healthcare) performed as per manufacturer's instructions. DNA samples from all patients were sent to Asper Ophthalmics (Tartu, Estonia) for analysis using the LCA chip [16]. This is a microarray which contained (at the time of the study) 344 disease associated SNPs and several common variant polymorphisms in six LCA associated genes and 2 early onset genes (*GUCY2D*, *CRX*, *RPE65*, *CRB1*, *RPGRIP1*, *AIPL1*, *LRAT*, and *MERTK*). Genorama genotyping software (Asper Ophthalmics Ltd) can subsequently produce a bidirectional allele call at each SNP being interrogated. The panel subsequently underwent direct sequencing of PCR products [11] performed using an ABI 3130 36 cm capillary sequencer (Applied Biosystems, Inc., Foster City, CA). 181 control DNA samples isolated from anonymous Aberdeen-based blood donors were screened for DNA mismatches using the Idaho Technology Inc. Light Scanner (Salt Lake City, UT). Fragments revealing a nucleotide mismatch were fully sequenced to identify the mutations.

RESULTS

A de novo heterozygous c.413C>G p.S138X stop mutation was identified in a seven-year-old male (Figure 1; Pt 13424) who was negative for all variants assayed by the LCA chip. The mutation was not present in either parent. This mutation was not detected in 181 controls. Known polymorphic single nucleotide polymorphisms (SNPs), were observed at similar frequencies in the disease cohort and the blood donor controls, indicating that the ethnic background of the patient cohort and the controls was comparable.

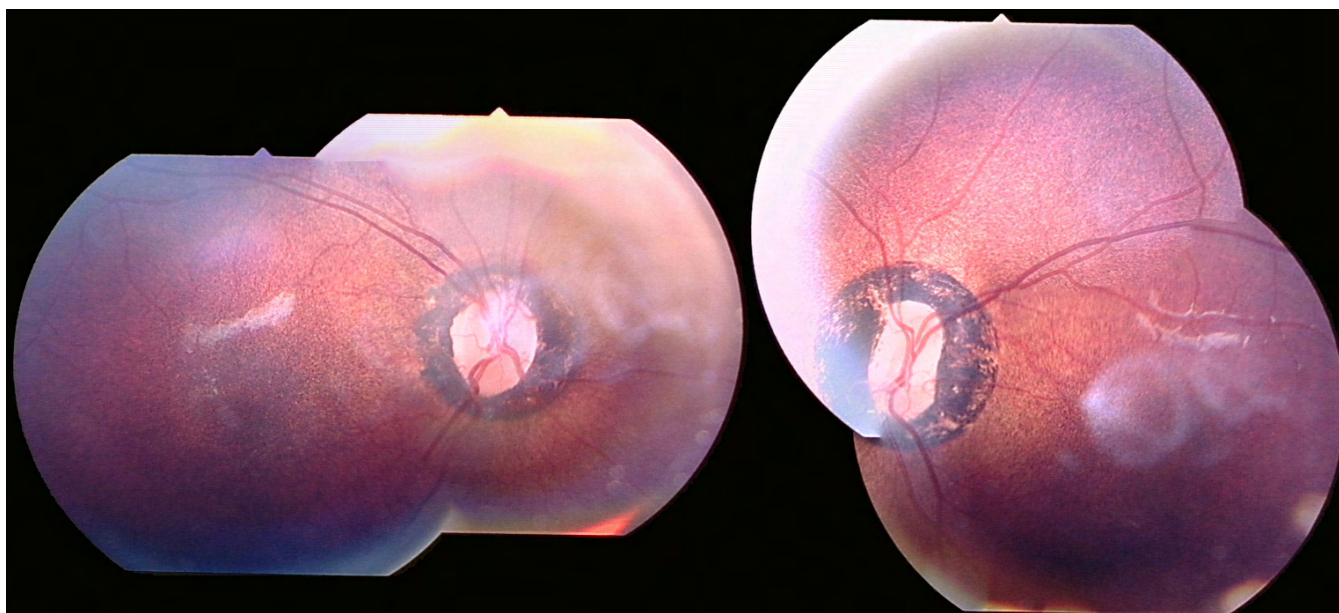


Figure 2. Fundus photography. Images are right and left eye color fundus photograph composites of patient NH13424 performed using TOPCON retinal camera at 2.45x magnification at 35 degrees. Composite images were created using Adobe Photoshop. Image cropping and reflections are observed secondary to poor pupil dilatation. The images show significant and abnormal peripapillary hyperpigmentation with fine granular pigmentation at the level of the retinal pigment epithelium (RPE).

The patient had a history of failure to thrive from birth and multiple investigations of the possible cause including investigations for Batten disease, Williams syndrome, and Russell-Silver dwarfism, all of which were negative. At 9 months old, required nasogastric feeding for 6 months following a history of failure to thrive. He has always been on the lowest percentiles for stature. Growth hormone deficiency was suggested indirectly via low levels of IGF1 and IGFBP3. An MRI scan was reported as normal. His parents noted poor vision and nyctalopia during his first year of life. There was no family history of eye disease. One younger sibling who has no visual problems was unavailable for further investigation.

Ophthalmic examination when he was six years old disclosed a small angle alternating esotropia with mild torsional nystagmus. Visual acuities were 0.4 and 0.5 logMAR in his right and left eyes, respectively. Cycloplegic retinoscopy revealed a refractive error of 0.00 diopter sphere/+3.00 diopter cylinder at 180 degrees in the right eye, and +3.00 diopter sphere/+2.50 diopter cylinder at 180 degrees in the left. He had normal color vision, according to Hardy Rand Rittler; he had normal pupil reactions and clear lenses. Funduscopy revealed a hyperpigmented area of retina surrounding the optic disc with a fine granular pigmentation of the RPE throughout the fundus (Figure 2). There was no vascular attenuation.

ERGs were performed according to international standards [13]. The dark-adapted bright flash ERG was markedly electronegative with additional a-wave reduction. Light-adapted flicker ERGs were delayed and subnormal and the single flash cone ERG was subnormal with a reduced b:a

ratio (Figure 3). The profoundly negative ERG suggests dysfunction that is both marked and at a level that is post-phototransduction, usually post-receptor. The a-wave was subnormal but not delayed, indicating additional loss of photoreceptor function. The ON-OFF and S-cone ERGs [17, 18] suggesting greater ON- than OFF- bipolar pathway involvement. The pattern ERG P50 component was reduced bilaterally, which is consistent with macular involvement. (Figure 3)

DISCUSSION

This study provides evidence of a novel mutation in *OTX2* associated with early onset retinal dystrophy and pituitary insufficiency. This mutation is predicted to lead a truncated protein. Chatelain et al. [19] observed that *OTX2* truncation at residues 106 and 161, flanking the site of the predicted termination, described here, leads to loss of transactivation function. This strongly suggests that the mutation identified in our patient is also likely to cause haploinsufficiency.

Clinical studies have demonstrated that mutations in *OTX2* are associated with a wide range of ocular phenotypes (Table 1) [7,11,20–22]. Experimentally, the role of *OTX2* in regional specification of the eye, particularly the RPE, has long been established from expression studies in normal and mutant animal models [1,4,23–26]. Subsequent studies have also demonstrated the role of *OTX2* in the development of both the retinal photoreceptors and bipolar cells [25,27]. It has been hypothesized that *OTX2* is involved in retinal photoreceptor cell fate but not in bipolar cell fate in early development, while it is involved in the terminal differentiation of both

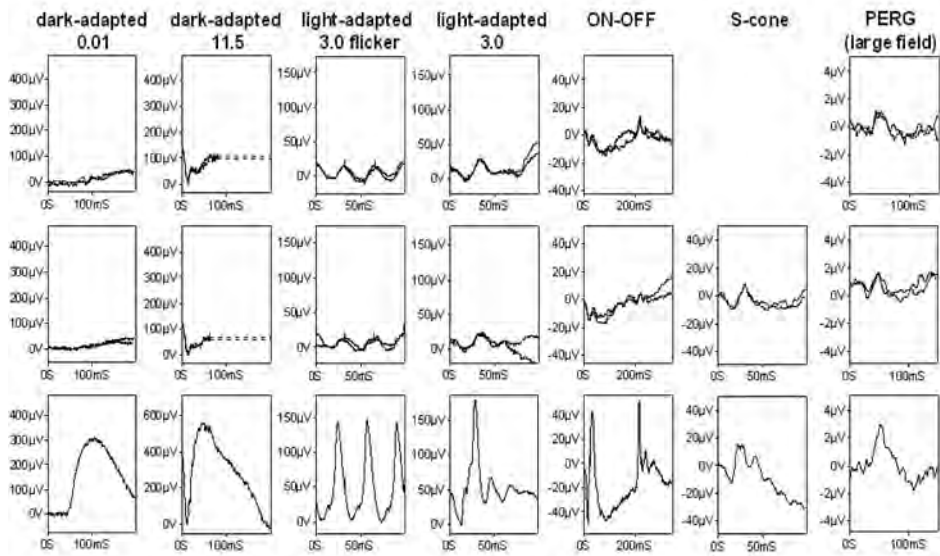


Figure 3. Electrophysiology. Full-field ERGs and pattern ERGs from the right (row 1) and left eye (row 2) of the patient and normal examples (row 3) for comparison. Full-field ERGs were performed using corneal electrodes in August 2007; pattern ERGs had been recorded to a 30 degree field checkerboard stimulus using periorbital surface electrodes two years earlier. Dark-adapted ERGs are shown for flash intensities of 0.01 and 11.5 cd.s.m⁻²; light-adapted ERGs for a flash intensity of 3.0 cd.s.m⁻² at 30 Hz (flicker) and at 2 Hz.. ON-OFF ERGs used an orange stimulus (560 cd.m⁻², duration 200 ms) superimposed on a green background (150 cd.m⁻²). S-cone ERGs used a blue stimulus (445 nm, 80 cd.m⁻²) on an orange background (620 nm, 560 cd.m⁻²). Eye movement artifacts are replaced by broken lines for clarity. S-cone ERGs were unavailable for the right eye.

TABLE 1. PREVIOUSLY PUBLISHED MUTATIONS IN *OTX2* WITH ASSOCIATED PHENOTYPES.

Nucleotide	Protein	Phenotype	Reference
c.674A>G	p.Asn225Ser	CPHD	[22]
c.674A>G	p.Asn225Ser	CPHD	[22]
whole gene deletion	whole gene deletion	Extreme microphthalmia	[20]
whole gene deletion	whole gene deletion	Anophthalmia	[20]
c.93C>G	p.Tyr31X	Microphthalmia	[20]
c.106dupC	p.Arg36ProfsX52	Microphthalmia OD/ normal OS	[20]
c.106dupC	p.Arg36ProfsX52	Anophthalmia OD/ Coloboma OS	[20]
c.289C>T	p.Gln97X	Extreme microphthalmia	[20]
c.289C>T	p.Gln97X	Inferior iris coloboma OD/ retinal coloboma OS	[20]
c.373_374delAG	p.Gly126TrpfsX11	Anophthalmia	[20]
c.404_405dupCT	p.Ser136LeufsX43	Anophthalmia and CPHD	[21]
c.402dupC	p.Ser135LeufsX2	Anophthalmia, GH deficiency; cleft palate	[7]
c.463_464dupGC	p.Ser156LeufsX23	bilateral anophthalmia; developmental delay	[11]
c.265C>G	p.Arg89Gly	bilateral microphthalmia	[11]
c.81delC	p.Ser28ProfsX23	bilateral severe microphthalmia	[11]
c.537T>A	p.Tyr179X	severe microphthalmia and colobomata, developmental delay seizures	[11]
c.537T>A	p.Tyr179X	bilateral mild microphthalmia, retinal dystrophy originally described as LCA	[11]
c.117_118delCC	p.Arg40GlyfsX47	bilateral anophthalmia	[11]
c.295C>T	p.Gln99X	bilateral anophthalmia	[11]
c.397C>A	p.Pro133Thr	bilateral microphthalmia	[11]
c.400C>G	p.Pro134Ala	anophthalmia OS/ OD normal	[11]

Mutations in *OTX2*. a summary of the published cases. Table lists previously published mutations in *OTX2* with associated phenotypes. The nucleotide and protein nomenclature used conforms to current standards and may differ from the notation used in the original publications (LOVD). Abbreviations: CPHD represents combined pituitary hormone deficiency; OD represents right eye; OS represents left eye; GH represents growth hormone.

photoreceptors and bipolar cells late in development in cooperation with *CRX* [8]. Pituitary dysfunction has been described in two patients with heterozygous 14q22–23 microdeletions involving *OTX2* [28,29]. Recently, the role of *OTX2* in pituitary development has been further delineated with three case reports of *OTX2* mutations associated with pituitary hormone deficiency (CPHD) [21,22]. A further case

of anophthalmia associated with an *OTX2* mutation was reported to have isolated growth hormone deficiency and short stature [7]. In the latter publication a requirement for *OTX2* binding was postulated at a site previously shown to lie in the promoter of *HESX1* [30].

The phenotypic spectrum described in this case report is consistent with the assigned multiple roles of *OTX2* in the development and function of both RPE and neural retinal, as well as in the pituitary. Mutations in *OTX2* are a rare cause of infantile onset retinal dystrophies and the finding of an electronegative ERG with additional a-wave reduction suggests that there is dysfunction at the level of the photoreceptor and also post-phototransduction, likely to involve both ON- and OFF- bipolar cells. Funduscopy revealed a fine granular pigmentation of the RPE. In addition, an unusual hyperpigmented juxtapapillary ring, not normally seen in retinal dystrophies, was observed. The history of early failure to thrive and subsequent short stature with low implied growth hormone levels adds further support for a role for *OTX2* in pituitary function.

ACKNOWLEDGMENTS

This work was supported by Foundation Fighting Blindness, Fight for Sight, and National Institute for Health Research Moorfields Eye Hospital Biomedical Research Centre.

REFERENCE LIST

1. Simeone A, Acampora D, Mallamaci A, Stornaiuolo A, D'Apice MR, Nigro V, Boncinelli E. A vertebrate gene related to orthodenticle contains a homeodomain of the bicoid class and demarcates anterior neuroectoderm in the gastrulating mouse embryo. *EMBO J* 1993; 12:2735-47. [PMID: 8101484]
2. Acampora D, Mazan S, Lallemand Y, Avantiaggiato V, Maury M, Simeone A, Brûlet P. Forebrain and midbrain regions are deleted in *Otx2*^{-/-} mutants due to a defective anterior neuroectoderm specification during gastrulation. *Development* 1995; 121:3279-90. [PMID: 7588062]
3. Ang SL, Jin O, Rhinn M, Daigle N, Stevenson L, Rossant J. A targeted mouse *Otx2* mutation leads to severe defects in gastrulation and formation of axial mesoderm and to deletion of rostral brain. *Development* 1996; 122:243-52. [PMID: 8565836]
4. Matsuo I, Kuratani S, Kimura C, Takeda N, Aizawa S. Mouse *Otx2* functions in the formation and patterning of rostral head. *Genes Dev* 1995; 9:2646-58. [PMID: 7590242]
5. Martínez-Morales JR, Dolez V, Rodrigo I, Zaccarini R, Leconte L, Bovolenta P, Saule S. *OTX2* activates the molecular network underlying retina pigment epithelium differentiation. *J Biol Chem* 2003; 278:21721-31. [PMID: 12663655]
6. Rath MF, Morin F, Shi Q, Klein DC, Moller M. Ontogenetic expression of the *Otx2* and *Crx* homeobox genes in the retina of the rat. *Exp Eye Res* 2007; 85:65-73. [PMID: 17467693]
7. Dateki S, Fukami M, Sato N, Muroya K, Adachi M, Ogata T. *OTX2* mutation in a patient with anophthalmia, short stature, and partial growth hormone deficiency: functional studies using the IRBP, HESX1, and POU1F1 promoters. *J Clin Endocrinol Metab* 2008; 93:3697-702. [PMID: 18628516]
8. Koike C, Nishida A, Ueno S, Saito H, Sanuki R, Sato S, Furukawa A, Aizawa S, Matsuo I, Suzuki N, Kondo M, Furukawa T. Functional roles of *Otx2* transcription factor in postnatal mouse retinal development. *Mol Cell Biol* 2007; 27:8318-29. [PMID: 17908793]
9. Sugiyama S, Prochiantz A, Hensch TK. From brain formation to plasticity: insights on *Otx2* homeoprotein. *Dev Growth Differ* 2009; 51:369-77. [PMID: 19298552]
10. Esumi N, Kachi S, Hackler L Jr, Masuda T, Yang Z, Campochiaro PA, Zack DJ. BEST1 expression in the retinal pigment epithelium is modulated by *OTX* family members. *Hum Mol Genet* 2009; 18:128-41. [PMID: 18849347]
11. Ragge NK, Brown AG, Poloschek CM, Lorenz B, Henderson RA, Clarke MP, Russell-Eggitt I, Fielder A, Gerrelli D, Martinez-Barbera JP, Ruddle P, Hurst J, Collin JR, Salt A, Cooper ST, Thompson PJ, Sisodiya SM, Williamson KA, Fitzpatrick DR, van Heyningen V, Hanson IM. Heterozygous mutations of *OTX2* cause severe ocular malformations. *Am J Hum Genet* 2005; 76:1008-22. [PMID: 15846561]
12. Holder GE, Brigell MG, Hawlina M, Meigen T, Vaegan, Bach M. ISCEV standard for clinical pattern electroretinography—2007 update. *Doc Ophthalmol* 2007; 114:111-6. [PMID: 17435967]
13. Marmor MF, Fulton AB, Holder GE, Miyake Y, Brigell M, Bach M. ISCEV Standard for full-field clinical electroretinography (2008 update). *Doc Ophthalmol* 2009; 118:69-77. [PMID: 19030905]
14. Holder GE, Robson AG. Paediatric Electrophysiology: A Practical Approach. In: Lorenz B, Moore AT, editors. *Paediatric Ophthalmology, Neuro-ophthalmology, Genetics*. Heidelberg: Springer; 2006: 133–55.
15. Kriss A, Thompson D. Visual Electrophysiology. In: Taylor D, editor. *Paediatric ophthalmology*. Oxford: Blackwell; 1997. p. 93–121.
16. Henderson RH, Waseem N, Searle R, van der Spuy J, Russell-Eggitt I, Bhattacharya SS, Thompson DA, Holder GE, Cheetham ME, Webster AR, Moore AT. An assessment of the apex microarray technology in genotyping patients with leber congenital amaurosis and early-onset severe retinal dystrophy. *Invest Ophthalmol Vis Sci* 2007; 48:5684-9. [PMID: 18055820]
17. Sieving PA. Photopic ON- and OFF-pathway abnormalities in retinal dystrophies. *Trans Am Ophthalmol Soc* 1993; 91:701-73. [PMID: 8140708]
18. Audo I, Michaelides M, Robson AG, Hawlina M, Vaclavik V, Sandbach JM, Neveu MM, Hogg CR, Hunt DM, Moore AT, Bird AC, Webster AR, Holder GE. Phenotypic variation in enhanced S-cone syndrome. *Invest Ophthalmol Vis Sci* 2008; 49:2082-93. [PMID: 18436841]
19. Chatelain G, Fossat N, Brun G, Lamonerie T. Molecular dissection reveals decreased activity and not dominant negative effect in human *OTX2* mutants. *J Mol Med* 2006; 84:604-15. [PMID: 16607563]
20. Wyatt A, Bakrania P, Bunyan DJ, Osborne RJ, Crolla JA, Salt A, Ayuso C, Newbury-Ecob R, Abou-Rayyah Y, Collin JR, Robinson D, Ragge N. Novel heterozygous *OTX2* mutations and whole gene deletions in anophthalmia, microphthalmia and coloboma. *Hum Mutat* 2008; 29:E278-83. [PMID: 18781617]
21. Tajima T, Ohtake A, Hoshino M, Amemiya S, Sasaki N, Ishizu K, Fujieda K. *OTX2* loss of function mutation causes anophthalmia and combined pituitary hormone deficiency with a small anterior and ectopic posterior pituitary. *J Clin Endocrinol Metab* 2009; 94:314-9. [PMID: 18854396]

22. Diaczok D, Romero C, Zunich J, Marshall I, Radovick S. A novel dominant negative mutation of OTX2 associated with combined pituitary hormone deficiency. *J Clin Endocrinol Metab* 2008; 93:4351-9. [PMID: 18728160]
23. Zuber ME, Gestri G, Viczian AS, Barsacchi G, Harris WA. Specification of the vertebrate eye by a network of eye field transcription factors. *Development* 2003; 130:5155-67. [PMID: 12944429]
24. Martinez-Morales JR, Signore M, Acampora D, Simeone A, Bovolenta P. Otx genes are required for tissue specification in the developing eye. *Development* 2001; 128:2019-30. [PMID: 11493524]
25. Larsen KB, Lutterodt M, Rath MF, Møller M. Expression of the homeobox genes PAX6, OTX2, and OTX1 in the early human fetal retina. *Int J Dev Neurosci* 2009; 27:485-92. [PMID: 19414065]
26. Hever AM, Williamson KA, van Heyningen V. Developmental malformations of the eye: the role of PAX6, SOX2 and OTX2. *Clin Genet* 2006; 69:459-70. [PMID: 16712695]
27. Kim DS, Matsuda T, Cepko CL. A core paired-type and POU homeodomain-containing transcription factor program drives retinal bipolar cell gene expression. *J Neurosci* 2008; 28:7748-64. [PMID: 18667607]
28. Elliott J, Maltby EL, Reynolds B. A case of deletion 14(q22.1→q22.3. associated with anophthalmia and pituitary abnormalities. *J Med Genet* 1993; 30:251-2. [PMID: 7682620]
29. Nolen LD, Amor D, Haywood A, St Heaps L, Willcock C, Mihelec M, Tam P, Billson F, Grigg J, Peters G, Jamieson RV. Deletion at 14q22–23 indicates a contiguous gene syndrome comprising anophthalmia, pituitary hypoplasia, and ear anomalies. *Am J Med Genet A* 2006; 140:1711-8. [PMID: 16835935]
30. Spieler D, Bäumer N, Stebler J, Köprunner M, Reichman-Fried M, Teichmann U, Raz E, Kessel M, Wittler L. Involvement of Pax6 and Otx2 in the forebrain-specific regulation of the vertebrate homeobox gene ANF/Hesx1. *Dev Biol* 2004; 269:567-79. [PMID: 15110720]